

California, Riverside, provided the financial support for this study.² A.M. also thanks the Spanish Ministry of Education and Sciences for a Postdoctoral fellowship.

Registry No.—1, 67-97-0; 7, 54473-74-4; 8, 64242-58-0; 9, 58028-00-5; **10a**, 26358-75-8; **10b**, 52032-63-0; **10c**, 56498-47-6; **10d**, 64265-01-6; **11a**, 64265-02-7; **11b**, 64252-59-1; **11c**, 64252-60-4; **12a**, 64252-61-5; **12b**, 64252-62-6; **13**, 64252-63-7; **14**, 53830-00-5; **15**, 64252-64-8; **16**, 52753-84-1; **17**, 21072-90-2; **18**, 5837-24-1; **19**, 21072-91-3; **20a**, 64265-03-8; **20b**, 64252-65-9; **21a**, 64252-66-0; **21b**, 64252-67-1; **21c**, 64265-09-4; **22a**, 35339-68-5; **22b**, 60008-81-3; **23**, 64252-54-6; **24a**, 64252-55-7; **24b**, 64252-56-8; **25**, 64252-57-9; **26a**, 50392-20-6; **26b**, 58699-24-4; **27b**, 58699-25-5; benzoyl chloride, 98-88-4; *p*-toluenesulfonylhydrazine, 1576-35-8.

References and Notes

- (1) (a) For a partial preliminary account, see W. H. Okamura and M. R. Pirio, *Tetrahedron Lett.*, 4317 (1975). (b) Paper 12 in this series: W. H. Okamura, M. L. Hammond, A. Rego, A. W. Norman, and R. M. Wing, *J. Org. Chem.*, **42**, 2284 (1977).
- (2) This study was supported by USPHS Grants AM-16595 and AM-9012 and by a grant from the Intramural Research Fund of the University of California, Riverside, Calif.
- (3) (a) Department of Chemistry; (b) Department of Biochemistry.
- (4) For exhaustive reviews on this subject, see (a) A. W. Norman and H. Henry, *Recent Prog. Horm. Res.*, **30**, 431 (1974); (b) J. L. Omdahl and H. F. DeLuca, *Physiol. Rev.*, **53**, 327 (1973); (c) also, the papers of E. Kodicek and co-workers, abundantly referenced in the two preceding review articles.
- (5) (a) E. V. Jensen and E. R. DeSombre, *Science*, **182**, 126 (1973); (b) B. W. O'Malley and A. R. Means, *ibid.*, **183**, 610 (1974); (c) H. H. Samuels and G. M. Tomkins, *J. Mol. Biol.*, **52**, 57 (1970).
- (6) (a) R. M. Wing, W. H. Okamura, M. R. Pirio, S. M. Sine, and A. W. Norman, *Science*, **186**, 939 (1974); (b) R. M. Wing, W. H. Okamura, A. Rego, M. R. Pirio, and A. W. Norman, *J. Am. Chem. Soc.*, **97**, 4980 (1975); (c) W. H. Okamura, M. L. Hammond, M. R. Pirio, R. M. Wing, A. Rego, M. N. Mitra, and A. W. Norman, "Proceedings of the Second Workshop on Vitamin D and Problems Related to Uremic Bone Disease", A. W. Norman, K. Schaefer, H. G. Grigoleit, and E. Ritz, Ed., Walter de Gruyter, Berlin, 1975, pp 259-278; (d) G. N. LaMar and D. L. Budd, *J. Am. Chem. Soc.*, **96**, 7317 (1974); (e) M. Sheves, E. Berman, D. Freeman, and Y. Mazur, *J. Chem. Soc., Chem. Commun.*, 643 (1975).
- (7) The equilibration of the A ring of vitamin D₃ between two chair forms was first discussed by Havinga and co-workers: (a) S. A. Bakker, J. Lugtenburg, and E. Havinga, *Recl. Trav. Chim. Pays-Bas*, **91**, 1459 (1972); (b) E. Havinga, *Experientia*, **29**, 1181 (1973).
- (8) W. H. Okamura, A. W. Norman, and R. M. Wing, *Proc. Natl. Acad. Sci. U.S.A.*, **71**, 4194 (1974).
- (9) See footnote 9, ref 6b.
- (10) W. H. Okamura, M. N. Mitra, D. A. Procsal, and A. W. Norman, *Biochem. Biophys. Res. Commun.*, **65**, 24 (1975).
- (11) (a) W. H. Okamura, M. N. Mitra, R. M. Wing, and A. W. Norman, *Biochem. Biophys. Res. Commun.*, **60**, 179 (1974); (b) M. N. Mitra, A. W. Norman, and W. H. Okamura, *J. Org. Chem.*, **39**, 2931 (1974); (c) A. W. Norman, M. N. Mitra, W. H. Okamura, and R. M. Wing, *Science*, **188**, 1013 (1975); H. Y. Lam, B. L. Onisko, H. K. Schnoes, and H. F. DeLuca, *Biochem. Biophys. Res. Commun.*, **59**, 845 (1974).
- (12) A. W. Norman, R. L. Johnson, T. W. Osborn, D. A. Procsal, S. C. Carey, M. L. Hammond, M. N. Mitra, M. R. Pirio, A. Rego, R. M. Wing, and W. H. Okamura, *Clin. Endocrinol. (Oxford)*, **5**, 121s (1976).
- (13) M. Ishiguro, A. Kajikawa, T. Haruyama, Y. Ogura, M. Okubayashi, M. Morisaki, and N. Ikekawa, *J. Chem. Soc., Perkin Trans. 1*, 2295 (1975).
- (14) A substituent at C₁ with an α configuration should be axial (or pseudoaxial), thus hindering nucleophilic attack at C₃ from the α face of the steroid. However, the principle of least motion, in the absence of a 1 α substituent, would argue for attack at C₃ from the α face (axial attack).
- (15) G. H. Posner and J.-S. Ting, *Tetrahedron Lett.*, 683 (1974).
- (16) C. R. Johnson and G. A. Dutra, *J. Am. Chem. Soc.*, **95**, 7777, 7783 (1973).
- (17) In one experiment, the C₁ acetate of **10b** was reacted with lithium dimethylcuprate. The product was identical with that obtained from the direct reaction with **10b**. It is not clear whether the acetate was removed during the reaction or during the workup. The directive influence by a hydroxyl toward an organometallic reagent has its analogy in the Simmons and Smith reaction. For one of several examples, see W. G. Dauben and A. C. Ashcraft, *J. Am. Chem. Soc.*, **85**, 3673 (1963).
- (18) In the pseudo-1 α -equatorial conformer, there is an acetoxy-11 α -H steric interaction, and the dienone is nonplanar (by distortion about the C₄-C₅ bond). In the pseudo-1 α -axial conformer, the dienone is planar, but there is a near-normal 1,3-diaxial interaction between the angular methyl group and the 2 β -H as well as one between the acetoxy and the 9 α -H. The pseudo-1 α -axial conformer should be the stabler of the pair. Consistent with this view is the observation that the 1 β -H appears as a broadened triplet of $J \sim 3$ Hz in its ¹H NMR spectrum.
- (19) (a) D. N. Jones, R. Grayshan, and D. E. Kime, *J. Chem. Soc. C*, 48 (1969); (b) D. N. Jones and D. E. Kime, *ibid.*, 846 (1966).
- (20) Similar differences have been observed in related olefins: R. L. Augustine and J. van Peppen, *Ann. N.Y. Acad. Sci.*, **158**, 482 (1969).
- (21) C. W. Shoppee and G. H. R. Summers, *J. Chem. Soc.*, 1786, 1790 (1952).
- (22) We are grateful to Professor G. H. Posner for an authentic specimen of 3 β -methylcholest-5-ene. (a) See ref 15; (b) R. H. Baker, L. S. Minckler, and O. R. Peterson, *J. Am. Chem. Soc.*, **77**, 3644 (1955).
- (23) By the method described in the Experimental Section for **12a**.
- (24) W. G. Dauben and D. S. Fullerton, *J. Org. Chem.*, **36**, 3277 (1971). See also R. M. Moriarty, H. Paaren, and J. Gilmore, *J. Chem. Soc., Chem. Commun.*, 927 (1974).
- (25) J. A. Marshall, R. A. Ruden, L. K. Hirsch, and M. Phillippe, *Tetrahedron Lett.*, 3795 (1971).
- (26) A. K. Bose, B. Lal, W. A. Hoffman, and M. S. Manhas, *Tetrahedron Lett.*, 1619 (1973).
- (27) H. H. Inhoffen and K. Irmischer, *Naturwissenschaften*, **45**, 86 (1958).
- (28) W. H. Okamura, M. L. Hammond, H. J. C. Jacobs, and J. V. Thuijl, *Tetrahedron Lett.*, 3807 (1976), and references cited therein.
- (29) F. A. L. Anet, *J. Am. Chem. Soc.*, **84**, 1053 (1962).
- (30) E. L. Eliel, N. L. Allinger, S. J. Angyal, and G. A. Morrison, "Conformational Analysis", Interscience, New York, N.Y., 1965, Chapters 1-2.
- (31) See p 52, ref 30.
- (32) (a) L. P. Kuhn, *J. Am. Chem. Soc.*, **74**, 2492 (1952); (b) H. Buc, *Ann. Chim. (Paris)*, **8**, 409 (1963).
- (33) We thank Professor Richard M. Wing and Dr. Albert Rego of this Department for the information.
- (34) We are grateful to Professor Yehuda Mazur for a preprint of a forthcoming publication which includes this information.
- (35) The method of assay has been described previously: K. Hibberd and A. W. Norman, *Biochem. Pharmacol.*, **18**, 2347 (1969).
- (36) Unpublished observations from this laboratory.
- (37) C. Djerrasi, R. R. Engle, and A. Bowers, *J. Org. Chem.*, **21**, 1547 (1956).
- (38) S. M. Sine, T. E. Conklin, and W. H. Okamura, *J. Org. Chem.*, **39**, 3797 (1974).
- (39) D. S. Fullerton and C. Chen, *Synth. Commun.*, **6**, 217 (1976).
- (40) H. O. House, C.-Y. Chu, J. M. Wilkins, and M. J. Umen, *J. Org. Chem.*, **40**, 1460 (1975).

Six New Bisbenzylisoquinoline Alkaloids from *Thalictrum rugosum*¹

Wu-Nan Wu, Jack L. Beal, Edward H. Fairchild, and Raymond W. Doskotch*

Division of Pharmacognosy and Natural Products Chemistry, College of Pharmacy,
Ohio State University, Columbus, Ohio 43210

Received July 26, 1977

The alkaloids thaligosidine (1), thaligosinine (11), thaligosine (14), thalirugine (19), thaliruginine (29), and thalirugidine (32) were isolated from the phenolic alkaloid fraction of *Thalictrum rugosum* Ait. roots. Their structures were advanced on the basis of spectral and chemical evidence.

The genus *Thalictrum* (family Ranunculaceae) has yielded well over 100 alkaloids biogenetically derivable from benzylisoquinoline precursors.² As part of a continuing study of alkaloids from *Thalictrum*, we report herein the isolation and structure determination of six new phenolic bisbenzyl-

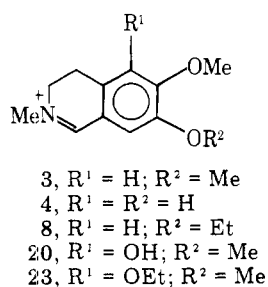
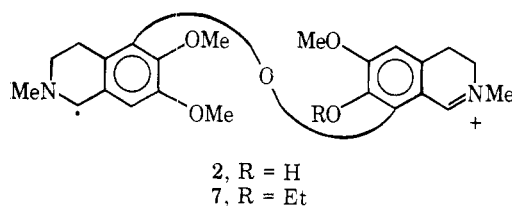
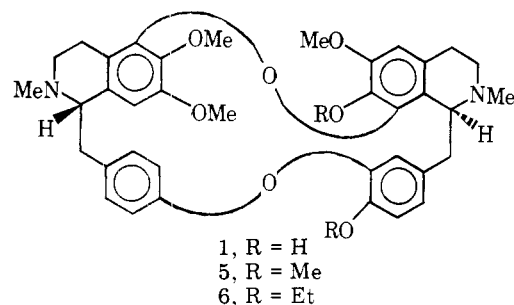
trahydroisoquinoline alkaloids from the roots of *Thalictrum rugosum* Ait. (*T. glaucum* Desf.). This source has already afforded over 20 alkaloids, of which seven have been characterized as bisbenzylisoquinolines.³

The residue obtained by extraction of the powdered roots

of *T. rugosum* was divided into the phenolic and nonphenolic tertiary alkaloid fractions by the usual solvent partition procedure, and the phenolic tertiary bases were chromatographed on silica gel and fractions thereof further purified to give the reported alkaloids.

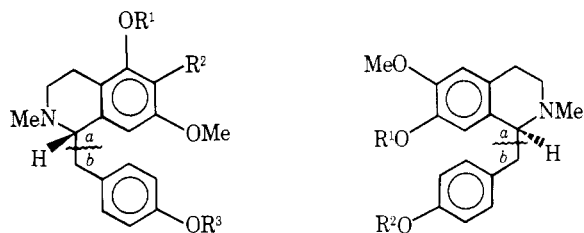
Thaligosidine (1), obtained as colorless crystals, mp 175–177 °C, was assigned the composition $C_{37}H_{40}N_2O_7$ by high-resolution mass spectrometry, and the 1H NMR spectrum clearly showed the presence of two *N*-methyls and three *O*-methyls along with nine aromatic protons. Two D_2O -exchangeable protons were assigned to two phenolic groups (vide infra). The UV spectrum exhibited a bathochromic shift in alkali. The IR spectrum showed hydroxyl absorption. The relatively intense molecular ion peak was suggestive of a bisbenzylisoquinoline structure with two diphenyl ethers,⁴ and the peak at m/e 412 for fragment 2 supported a head to head arrangement as well as the placement of one phenolic group in an isoquinoline ring. Additional fragment peaks assigned to ions 3 and 4 supported the corresponding partial structures.

Methylation of thaligosidine (1) with diazomethane gave a methylated derivative 5 that showed two additional methoxy

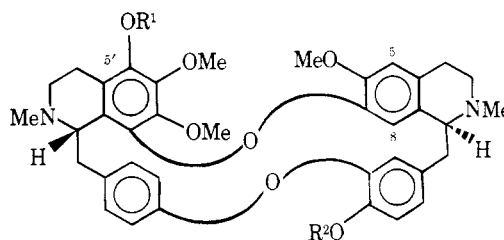


peaks in the 1H NMR spectrum and was identical with thalidasine (*S,S* configuration), previously isolated from this plant.⁵ This conversion established the carbon skeleton, oxygenation pattern, and stereochemistry for thaligosidine (1). The location of the phenolic groups was determined from studies on *O,O*-diethylthaligosidine (6). The mass spectral peaks at m/e 440 and 220 for fragments 7 and 8, respectively, confirmed the location of one phenolic group in an isoquinoline unit, and sodium–ammonia cleavage of 6 produced (*S*)-*N*-methyl-*O,O*-diethylcoclaurine (9)⁶ and (*S*)-5-hydroxyar-mepavine (10).⁵ The nonphenolic product established the positions of the two phenolic groups.

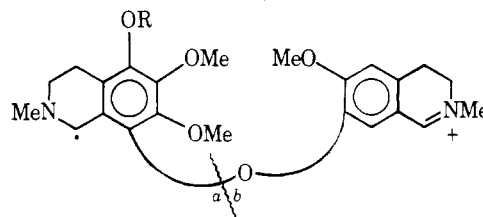
Thaligosinine (11), as colorless crystals, mp 233–234.5 °C (dec), with composition $C_{38}H_{42}N_2O_7$ from high-resolution mass measurement showed in the 1H NMR spectrum two *N*-methyls, four *O*-methyls, and nine aromatic protons. A phenolic hydroxyl was supported by absorption in the IR and NMR spectra (in the latter, D_2O exchangeable), and by ob-



- 10, R¹ = R³ = H; R² = OMe
18, R¹ = Et; R² = OMe; R³ = H
35, R¹ = Et; R² = OMe, R³ = Me
36, R¹ = Et; R² = H; R³ = Me
- 9, R¹ = R² = Et
17, R¹ = H; R² = Me
24, R¹ = Et; R² = Me
31, R¹ = R² = Me



- 11, R¹ = Me; R² = H
12, R¹ = R² = Me
14, R¹ = H; R² = Me
16, R¹ = Et; R² = Me



- 13, R = Me
15, R = H

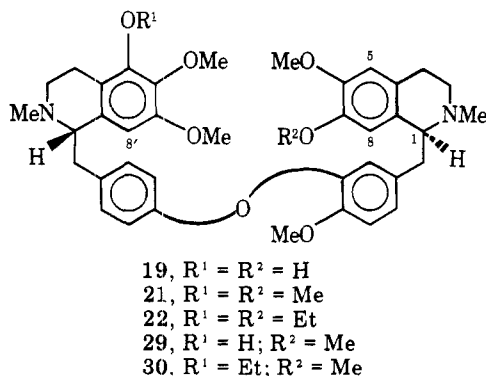
servation of a bathochromic shift in the UV spectrum with alkali. The *O*-methyl derivative was identical with thalrugosaminine (12) (*S,S* configuration) and established for thaligosinine the complete structure except for the phenolic hydroxyl position. Since the MS of thaligosinine (11) contains a peak at m/e 426 corresponding to fragment 13 and other peaks consistent with 13, the phenolic hydroxyl must be with the benzylic ring.

Thaligosine (14), mp 143–145 °C, was found to have the same elemental composition as thaligosinine (11) and the NMR spectrum appeared similar; the major difference was in the location of one of the methoxys. On treatment with diazomethane, thaligosine (14) was converted to thalrugosaminine (12), making thaligosine (14) a position isomer of thaligosinine (11). The phenolic group must be located in the tetraoxygenated isoquinoline ring, since the MS spectrum of 14 contains peaks at m/e 412, 222, and 192 corresponding to fragments 15, 15a + H and 15b + H, respectively. *O*-Ethylthaligosine (16) on sodium–ammonia cleavage afforded two phenolic products 17 and 18. The former is identical with a cleavage product from thalrugosaminine (12).^{3a} The latter is new, for which the physical data (NMR, MS) support a structure in which the isoquinoline ring bears an ethoxy and two methoxy groups, and the benzylic ring contains a para phenolic hydroxyl. This compound is identical to one of the degradation products from thalirugine (19) for which location of the ethylated phenol was established (vide infra). Thaligosine (14) therefore has the phenolic group at the 5'-position and configuration *S,S*.

Thalirugine (19) was isolated as an amorphous solid and assigned a dimeric benzylisoquinoline structure on the basis of the number of protons in the NMR spectrum. The mass spectrum showed only very weak peaks above m/e 400 but the elemental analyses were consistent with the formula

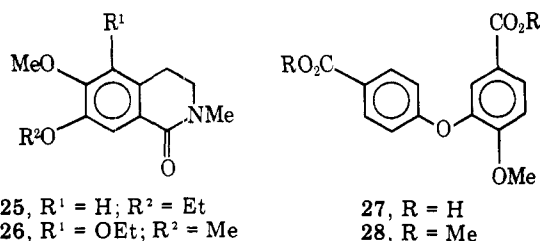
$C_{39}H_{44}N_2O_7$ for which an apparent molecular ion at m/e 640 of 0.01% intensity was present. Such weak molecular peaks are observed for single diphenyl ether linked dimeric alkaloids.⁴ The NMR spectrum contained peaks for two *N*-methyls, four *O*-methyls, ten aromatic protons, and two phenolic groups. Positive phosphomolybdic acid⁷ and Gibbs⁸ tests supported a para unsubstituted phenol. Partial structures for the two isoquinoline portions in which one contains a phenolic and two methoxyl groups and the other a phenolic and one methoxy were supported by ions in the mass spectrum at m/e 220 and 192 for fragments 20 and 4 (or their equivalent), respectively.

Methylation of thalirugine (19) with diazomethane gave a dimethyl derivative 21, which substantiated the presence of



two phenolic hydroxyls and their location in the isoquinoline rings, as did the diethyl derivative 22. For the latter compound the intense mass spectral peaks at m/e 250 and 220 correspond to fragment ions 23 and 8, respectively. Sodium in ammonia cleavage of *O,O*-diethylthalirugine (22) yielded a nonphenolic base 24 that was identical with a cleavage product from *O*-ethylthalirugosidine;⁵ thus, one of the phenolic groups in thalirugine (19) is at position 7, and the configuration at C-1 *S*. The phenolic cleavage product was found to be identical with product 18 from *O*-ethylthaligosine (16); therefore, the other asymmetric center of thalirugine (19) is also *S*. The two cleavage products identified the nature of the oxygenation pattern for the individual benzylisoquinoline units of thalirugine (19). In addition, the phenolic group in the isoquinoline ring of 18 must be placed at position 5 if thalirugine is to give a positive Gibbs' test. It follows that thaligosine (14) must bear the phenolic hydroxyl at 5'.

Permanganate oxidation of *O,O*-diethylthalirugine (22) afforded three products, two isoquinolones 25^{9,10} and 26 and

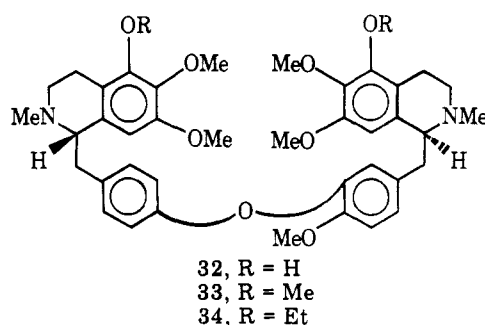


diphenyl ether dicarboxylic acid 27, each identified by direct comparison with known samples. 2-Methyl-5-ethoxy-6,7-dimethoxy-1-oxo-1,2,3,4-tetrahydroisoquinoline (26) was prepared by oxidation of the diethyl ether product of 10, a sodium-ammonia cleavage product from thalidasine (5).¹¹ The isoquinolones confirmed the phenol locations in the isoquinoline rings, and the diphenyl ether, also characterized as the diester 28, established the point of attachment for the monomeric units of thalirugine (19).

Thalirugine (29) was obtained as an amorphous solid, $[\alpha]_D +104^\circ$, and exhibited spectral characteristics of a bisbenzylisoquinoline alkaloid with one diphenyl ether structure (weak

molecular ion in MS). The NMR spectrum showed peaks for two *N*-methyls, five *O*-methyls, and ten aromatic protons. Treatment of thalirugine (29) with diazomethane formed a monomethyl derivative 21 identical with the methylated product from thalirugine (19); thus, thalirugine (29) is one of two *O*-methylthalirugines, and its MS with a peak at m/e 222 (20) supported a phenolic group in the trioxxygenated isoquinoline ring. The positive Gibbs' test would place the phenolic group at position 5'. Confirmation of this assignment was made by the isolation of benzyl isoquinoline 18 on sodium-ammonium cleavage of *O*-ethylthalirugine (30). The other cleavage product was (*S*)-*O*-methylarmepavine (31).

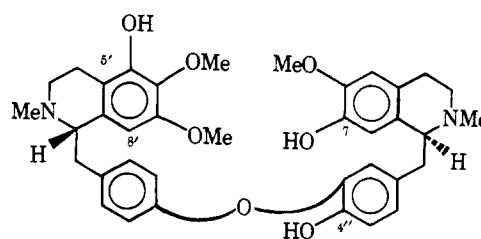
Thalirugidine (32), the third monoether-linked bisalkaloid, was assigned formula $C_{39}H_{46}N_2O_8$ on the basis of MS and elemental analyses. The NMR spectrum contained peaks for two *N*-methyls, five *O*-methyls, and nine aromatic protons, two of which were located at upfield positions, characteristic of H-8 and H-8' protons. Two D_2O -exchangeable protons were from phenols and were supported by the preparation of di-*O*-methyl 33 and di-*O*-ethyl 34 derivatives. The MS of



thalirugidine (32) and its derivatives, all with weak M^+ peaks, were in agreement with the location of a phenolic hydroxyl on each of the two trioxxygenated isoquinoline rings. For example, peaks are present at m/e 222 for fragment 20 and m/e 250 for 23 of thalirugidine (32) and diethyl ether 34, respectively.

Sodium-ammonia reductive cleavage of 34 gave (*S*)-5-ethoxy-4',6,7-trimethoxy-*N*-methyltetrahydroisoquinoline (35) as the nonphenolic product and (*S*)-5-ethoxy-4'-hydroxy-6,7-dimethoxy-*N*-methyltetrahydroisoquinoline (18) as the phenolic product. The configuration was assigned on the basis of positive CD maxima at 282 and 230 nm for cleavage products 18 and 35. Methylation converted 18 to 35. Permanganate oxidation of *O,O*-diethylthalirugidine (34) formed 2-methoxy-4',5-dicarboxy diphenyl ether (27) (identified as the dimethyl ester 28) and provided the evidence for the location of the ether linkage. In addition, the isoquinolone 26 was also isolated. The CD spectrum of thalirugidine (32), with two positive maxima at 280 and 230 nm, is consistent with the *S,S* stereochemistry.

Five of the six new alkaloids, in this report, are derivable biogenetically from a hypothetical monoether-linked bisalkaloid 37 (northalirugine) formed by phenol coupling of (*S*)-northalifendlerine and (*S*)-*N*-methylcoclaurine. Intramolecular phenol coupling of 37 between the 5'-OH and the 8 position would yield thaligosidine (1), which on methylation



of both phenolic groups forms thalidasine (5). With phenol coupling between the 7-OH and the 8' position, the precursor of thaligosinine (11) and thaligosine (14) would result. Only methylation, respectively, of the 5'-OH and the 4''-OH is required to the final products. Complete methylation gives thalrugosaminine (12). The remaining two alkaloids, thalirugine (19) and thaliruginine (29), are simple mono- and dimethylated products of the hypothetical precursor 37. The sixth alkaloid, thalirugidine (32), is a monomethylated product of another hypothetical precursor formed from two (S)-nortalifendlerines.

Experimental Section

Melting points are uncorrected. NMR spectra were determined in stated solvents with Me₄Si as an internal standard using Varian A-60A or Bruker HX-90E instruments and with chemical shifts (δ) reported in ppm and coupling constants (J) in Hz. IR and UV spectra were taken in CHCl₃ on a Beckman IR 4230 and in MeOH on a Cary 15 instrument, respectively. Mass spectra were obtained on an AEI MS-9 or DuPont 21-491 instrument by direct inlet probe at 70 eV. Optical rotations were measured on a Perkin-Elmer 241 photoelectric polarimeter and CD spectra in MeOH on a Durrum-Jasco ORD/UV-5 spectropolarimeter with Sproul Scientific SS-20 modification. TLC was performed on silica gel G (E. Merck) and column chromatography on silica gel PF254 (Brinkmann-EM) or alumina (Woelm) with stated solvents. Detection on TLC was with Dragendorff's spray reagent. Microanalyses were by Scandinavian Microanalytical Laboratory, Herlev, Denmark. Roots of *T. rugosum* were collected from plants grown in the College of Pharmacy Medicinal Plant Garden. A voucher specimen is on file.

Extraction of *T. rugosum* Roots and Initial Partitioning. Ground roots (17.7 kg) were percolated to exhaustion with ethanol, and the extract was evaporated to dryness at reduced pressure and 40 °C, leaving 1.5 kg of residue. Suspension of the residue in 18 L of 2% aqueous citric acid and filtering gave a filtrate that was extracted with an equal volume of CHCl₃ and then brought to pH 9.0 with NH₄OH. The basic solution was extracted successively with Et₂O (35 L) and CHCl₃ (18 L), and the Et₂O fraction was extracted with 12 L of 5% NaOH. Evaporation of the Et₂O left 75 g of nonphenolic tertiary alkaloids, while from the CHCl₃ extract 11.6 g of alkaloidal residue remained. The NaOH solution was treated with solid NH₄Cl and the cloudy suspension was extracted with 30 L of Et₂O from which 39.2 g of phenolic tertiary alkaloids was obtained.

Chromatography of the Phenolic Tertiary Alkaloids. The crude phenolic tertiary alkaloid fraction (30 g) was separated on a column of silica gel (0.9 kg), collecting 0.5-L fractions for analysis. The eluting solvents were CHCl₃ (5 L) and the following mixtures of MeOH in CHCl₃: 2.5% (10 L), 5% (12 L), 7.5% (8 L), 10% (6 L), 15% (3 L), 20% (6 L), 40% (4 L), and 50% (4 L). Final column washes were made with 4 L of MeOH and 2 L of 5% HOAc in MeOH. The eluted fractions gave the alkaloids that follow.

Thaligosidine (1). The residue (1.1 g) from fractions 28–31 was rechromatographed on 35 g of silica gel. The 1% MeOH in CHCl₃ eluates gave 230 mg of a residue that was crystallized from benzene to yield 180 mg of colorless crystalline thaligosidine (1): mp 175–177 °C; R_f 0.9 on TLC with PhH–Me₂CO–NH₄OH (20:20:0.5); $[\alpha]_D^{20}$ –45° (c 0.26, MeOH); CD $[\theta]_{287}$ –19 000, $[\theta]_{268}$ +6700, $[\theta]_{242}$ +5490 and $[\theta]_{224}$ –31 800; UV λ_{max} 275 (log ϵ 3.72), 283 nm (3.72), and in 0.01 N NaOH 275 (3.75), 284 (3.76), and 310 nm (shld) (3.22); IR (CHCl₃) 3540 cm⁻¹ (OH); NMR δ (CDCl₃) 2.25 and 2.66 (s, 2 NMe), 3.49, 3.75 and 3.86 (s, 3 OMe), 6.2–7.7 (m, 9 ArH), and 5.6 ppm (br, 2 OH, D₂O exchangeable), with benzene adduct showing peaks at δ (CDCl₃) 2.42 and 2.66 (s, 2 NMe), 3.43, 3.76, 3.85 (s, 3 OMe), 6.3–7.7 (m, 9 ArH), and 5.88 ppm (br, 2 OH); MS m/e 624.2849 (40%, M⁺, C₃₇H₄₀N₂O₇ requires 624.2835), 412 (7, 2), 411 (20, 2 – H), 206 (23, double ions of 412 and 3), and 192 (100, 4).

Anal. Calcd for C₃₇H₄₀N₂O₇· $\frac{3}{2}$ H₂O: C, 68.24; H, 6.18; N, 4.30. Found: C, 68.50; H, 6.48; N, 3.81.

O-Methylation of Thaligosidine (1). To 35 mg of thaligosidine (1) in 2.5 mL of MeOH was added ethereal diazomethane prepared from 1 g of *N*-methyl-*N*-nitroso-*p*-toluenesulfonamide. After 4 days, the residue remaining after evaporation of solvent was chromatographed on neutral alumina with PhH–CHCl₃ (1:1) to give 25 mg of a pale-yellow amorphous base showing identical properties (IR, UV, specific rotation, CD, NMR, and TLC) with authentic thalidasine (5) previously isolated from *T. rugosum*.⁵

O,O-Diethylthaligosidine (6). To 140 mg of thaligosidine (1) in 10 mL of MeOH was added ethereal diazoethane generated from 2

g of *N*-ethyl-*N'*-nitro-*N*-nitrosoguanidine. After 5 days the reaction residue was chromatographed on neutral alumina with benzene as solvent. The crystalline residue on recrystallization from MeOH gave colorless needles: mp 198–200 °C; R_f 0.93 on TLC with PhH–Me₂CO–NH₄OH (20:20:0.5); NMR δ (CDCl₃) 0.74 and 1.43 (2 t, J = 7, 2 OCH₂CH₃), 4.14 (q, J = 7, OCH₂CH₃) with another methylene quartet of ethoxy hidden in the methylene region, 2.24 and 2.62 (s, 2 NMe), 3.52, 3.74, and 3.87 (s, 3 OMe), and 6.2–7.6 (m, 9 ArH); MS (CI, isobutane) m/e 681 (100%, MH⁺, C₄₁H₄₉N₂O₇ requires 681), EI m/e 680 (100%, M⁺), 651 (11, M – Et), 440 (7, 7), 439 (22, 7 – H), 220 (37, 8), 206 (4, 3) and 190 (10, 8 – EtH).

Anal. Calcd for C₄₁H₄₉N₂O₇: C, 72.33; H, 7.11; N, 4.12. Found: C, 71.94; H, 7.15; N, 4.09.

Na/NH₃ Cleavage of O,O-Diethylthaligosidine (6). To 20 mL of liquid NH₃ containing 150 mg of Na at below –50 °C was added dropwise in 0.5 h 8 mL of tetrahydrofuran solution containing 70 mg of 6. The reaction was maintained for 2 h. The NH₃ was allowed to evaporate and the unreacted Na was decomposed with excess MeOH. The mixture was concentrated in vacuo to 2 mL, taken up in 250 mL of Et₂O, and extracted with 125 mL of 1 N NaOH to separate the products into phenolic and nonphenolic fractions.

From the washed (H₂O) and dried (Na₂SO₄) Et₂O extract was obtained 30 mg of an oil on evaporation that showed one spot, R_f 0.80, on TLC with PhH–Me₂CO–NH₄OH (20:20:0.6). Column chromatography on 2 g of neutral alumina with 100 mL of PhH–CHCl₃ (1:1) and 150 mL of CHCl₃ as eluents yielded 24 mg of 9 as an amorphous solid: NMR δ (CDCl₃) 1.31 and 1.38 (2 t, J = 7, 2 OCH₂CH₃), 3.76 and 3.98 (2 q, J = 7, 2 OCH₂CH₃), 2.51 (s, NMe), 3.82 (s, OMe), 6.09 (s, H-8), 6.55 (s, H-5), and 6.77 and 6.98 (AA'BB' q, J_{AB} = 8.5, 4 ArH); MS (CI, isobutane) m/e 356 (75%, MH⁺, C₂₂H₂₉NO₃ requires 355), 220 (100, a) and 135 (5, b); CD $[\theta]_{290}$ +9100, $[\theta]_{272}$ –1700, and $[\theta]_{234}$ +47 000. This compound showed identical TLC behavior, UV, IR, and NMR spectra with that of the corresponding cleavage product prepared from *O*,*O*-diethyllobamegine,⁶ but the CD spectrum was antipodal ($[\theta]_{290}$ –9300, $[\theta]_{272}$ +1900, and $[\theta]_{234}$ –38 000).

The NaOH solution was treated with NH₄Cl to pH 8–9 and extracted with Et₂O. The washed (H₂O) and dried (Na₂SO₄) Et₂O solution on evaporation gave 19 mg of a residue that showed two spots on TLC, R_f 0.6 (major) and 0.7, with PhH–Me₂CO–NH₄OH (20:20:0.6). Chromatography on silica gel (2 g) with CHCl₃, 2% and 4% MeOH in CHCl₃, yielded 12 mg of 10, identical (TLC, UV, IR, NMR, MS, and CD) with the cleavage product from *O*-ethylthalrugosidine.⁵

Thaligosinine (11). The column fractions 28–31 that on rechromatography afforded thaligosidine (1) gave on elution with 2% MeOH in CHCl₃ 526 mg of an early fraction that crystallized from Et₂O to give white crystals of thalrugosine, mp 206–207 °C, already reported from this source.⁵ The latter 2% MeOH in CHCl₃ effluents yielded 35 mg of a white crystalline solid that crystallized from Et₂O to give thaligosinine (11): mp 233–234.5 °C (dec); $[\alpha]_D^{21}$ –58.5° (c 0.316, MeOH); CD $[\theta]_{275}$ –13 000, $[\theta]_{242}$ –48 000, $[\theta]_{230}$ +100 000; UV λ_{max} 282 nm (log ϵ 3.90) and in 0.01 N NaOH 284 (4.11) and 307 nm (shld) (3.53); IR (CHCl₃) 3540 cm⁻¹; NMR δ (CDCl₃, 90 MHz) 2.51 and 2.56 (2 s, 2 NMe), 3.04, 3.40, 3.80, and 3.84 (4 s, 4 OMe), 6.36 (s, H-8), 6.47 (s, H-5), 6.5–7.5 (m, 7 ArH), and ~5.0 (br, OH, D₂O exchangeable); MS 638.3005 (100%, M⁺, C₃₈H₄₂N₂O₇ requires 638.2992), 426 (15, 13), 425 (37, 13 – H), 411 (34, 13 – Me), 236 (2, 13a), 213 (97, double ion of 13), 192 (31, 13b + H), 191 (6, 13b), and 190 (9, 13b – H).

Methylation of Thaligosinine (11) to Thalrugosaminine (12). A 25-mg sample of 11 in 3 mL of MeOH was treated with excess ethereal diazomethane for 3 days. The product was chromatographed on 2 g of neutral alumina with PhH–CHCl₃ (1:1) to give 23 mg of a pale-yellow solid. Its TLC mobility and UV, IR, NMR, and CD spectra were identical with those of thalrugosaminine (12) earlier reported from this plant.^{3a}

Thaligosine (14). The residue (1 g) from column fractions 32–34 was rechromatographed on 50 g of silica gel with CHCl₃ and 1, 2, 3, and 4% MeOH in CHCl₃. The 2 and 3% MeOH in CHCl₃ effluents gave a residue that from Et₂O gave 210 mg of thaligosine (14) as colorless crystals: mp 143–145 °C; R_f 0.33 on TLC with PhH–Me₂CO–NH₄OH (20:20:0.8); $[\alpha]_D$ –109° (c 0.17, MeOH); CD $[\theta]_{287}$ +3200, $[\theta]_{272}$ –1200, $[\theta]_{240}$ –7980, $[\theta]_{225}$ +16 700; UV λ_{max} 282 nm (log ϵ 3.86) and in 0.01 N NaOH 282 (3.88) and 305 nm (shld) (3.64); IR (CHCl₃) 3520 cm⁻¹ (OH); NMR δ (CDCl₃) 2.52 and 2.56 (2 s, 2 NMe), 3.08, 3.39, 3.78, and 3.95 (4 s, 4 OMe), 6.38 (s, H-8), 6.46 (s, H-5), 6.6–7.4 (m, 7 ArH) and 4.7 (br, OH, D₂O exchangeable); MS m/e 638 (89%, M⁺, C₃₈H₄₂N₂O₇ requires 638), 637 (27, M – H), 623 (4, M – Me), 412 (25, 15), 411 (92, 15 – H), 222 (14, 15a + H), 206 (100, double ion of 15) and 192 (30, 15b + H).

Anal. Calcd for C₃₈H₄₂N₂O₇·H₂O: C, 69.49; H, 6.75; N, 4.27. Found: C, 69.35; H, 6.60; N, 4.17.

O-Methylation of Thaligosine (14). A 30-mg sample of 14 in 5 mL of MeOH was treated with excess ethereal diazomethane for 4 days. The product was chromatographed on 1.5 g of neutral alumina with 100 mL of chloroform as eluent and the homogeneous [TLC, R_f 0.5 with PhH-Me₂CO-NH₄OH (20:20:0.8)] material (23 mg) was identical (UV, IR, MS, NMR, and CD) with thalugosamine (12).^{3a}

O-Ethylthalogosine (16). A 120-mg sample of thaligosine (14) in 5 mL of MeOH was treated with ethereal diazoethane prepared from 1.5 g of *N*-ethyl-*N'*-nitro-*N*-nitrosoguanidine. After 3 days the solvent was evaporated and the residue chromatographed on 6 g of neutral alumina with PhH-CHCl₃ (1:1) and CHCl₃ as eluents. The ethyl ether 16 as a homogeneous solid (110 mg) with R_f 0.47 on TLC with PhH-Me₂CO-NH₄OH (20:20:0.6) was eluted with the latter solvent and showed: NMR δ (CDCl₃) 1.38 (t, $J = 7$, OCH₂CH₃), ~4.2 (q, $J = 7$, OCH₂CH₃, partially obscured), 2.50 and 2.55 (2 s, 2 NMe), 3.08, 3.38, 3.78, and 3.92 (4 s, 4 OMe), 6.39 (s, H-8), 6.45 (s, H-5), and 6.6-7.5 (m, 7 ArH); MS m/e 666.3316 (100%, M⁺, C₄₀H₄₆N₂O₇ requires 666.3305).

Anal. Calcd for C₄₀H₄₆N₂O₇·0.5H₂O: C, 71.09; H, 7.01; N, 4.15. Found: C, 71.45; H, 7.03; N, 4.14.

Na/NH₃ Cleavage of O-Ethylthalogosine (16). The ethyl ether 16 (105 mg) was dissolved in 6 mL of tetrahydrofuran and reduced with Na in NH₃ as described for 6. The reaction mixture was separated into phenolic and nonphenolic fractions. Only unreacted starting material (5 mg) was obtained from the nonphenolic fraction. The phenolic fraction (57 mg) showed two major components on TLC, R_f 0.58 and 0.36 with PhH-Me₂CO-NH₄OH (20:20:0.6), and was separated on 3 g of silica gel.

The 1% MeOH in CHCl₃ eluent gave 13 mg of 17 as a yellow solid, identical (UV, IR, NMR, and CD) with one of the cleavage products from thalugosamine (12).^{3a}

The 2% MeOH in CHCl₃ eluent gave 20 mg of 18 which crystallized from MeOH: mp 113-114 °C; $[\alpha]_D^{21} +103$ (c 0.24, MeOH); CD $[\theta]_{284} +7300$, $[\theta]_{230} +56800$; UV λ_{max} 283 nm (log ϵ 3.61); IR (CHCl₃) 3595 cm⁻¹; NMR δ (CDCl₃) 1.34 (t, $J = 7$, OCH₂CH₃), 4.05 (q, $J = 7$, OCH₂CH₃), 2.50 (s, NMe), 3.54 and 3.82 (2 s, 2 OMe), 5.87 (s, H-8), 6.62 and 6.88 (AA'BB' q, $J_{AB} = 9$, benzylic ring H), and 6.45 (br, OH, lost in D₂O); MS m/e 357.1946 (0.5%, M⁺, C₂₁H₂₇NO₄ requires 357.1940, CI (isobutane) m/e 358 (10, MH⁺), 250 (100, a), 107 (2, b).

Thalirugine (19). The residue (1.9 g) from fractions 38-44 was rechromatographed on 200 g of neutral alumina with CHCl₃ as eluent. The residue precipitated from Et₂O to give thalirugine (19) as a white amorphous solid: R_f 0.52 on TLC with PhH-Me₂CO-NH₄OH (20:20:0.8); $[\alpha]_D^{20} +92^\circ$ (c 0.25, MeOH); CD $[\theta]_{282} +6400$, $[\theta]_{248} -3100$, and $[\theta]_{226} +78000$; UV λ_{max} 280 nm (log ϵ 3.81) and in 0.01 N NaOH 280 (3.86), 285 (3.87), and 310 nm (shld) (3.45); IR (CHCl₃) 3520 cm⁻¹ (OH); NMR (CDCl₃) δ 2.43 and 2.49 (2 s, 2 NMe), 3.58 and 3.83 (2 s, 2 OMe), 3.78 (s, 2 OMe), 5.73 (s, H-8'), 6.38 (s, H-8), 6.47 (s, H-5), 6.6-7.2 (m, 7 ArH), and 5.50 (br, OH, D₂O exchangeable); MS m/e 640 (0.01%, M⁺), 222 (100, 20), 207 (35, 20 - Me), and 192 (83, 4). Positive tests were obtained with phosphomolybdic acid⁷ and Gibbs' reagents.⁸

Anal. Calcd for C₃₈H₄₄N₂O₇·0.5H₂O: C, 70.24; H, 6.98; N, 4.31. Found: C, 70.11; H, 6.96; N, 4.27.

O-Methylation of Thalirugine (19) to 21. A 35-mg sample of 19 in 3 mL of MeOH was treated with excess ethereal diazomethane for 3 days. The residue after evaporation of solvent was chromatographed on 2 g of neutral alumina with CHCl₃ to yield derivative 21 as a pale yellow amorphous solid: R_f 0.75 on TLC with PhH-Me₂CO-NH₄OH (20:20:0.8); $[\alpha]_D^{21} +105^\circ$ (c 0.21, MeOH); CD $[\theta]_{282} +14700$, $[\theta]_{228} +76600$; NMR (CDCl₃) δ 2.48 and 2.51 (2 s, 2 NMe), 3.58, 3.63, 3.79, and 3.82 (4 s, 4 OMe), and 3.84 (s, 2 OMe), 5.93 (s, H-8'), 6.13 (s, H-8), 6.53 (s, H-5), and 6.6-7.2 (m, 7 ArH); MS m/e 668 (0.3%, M⁺, C₄₀H₄₈N₂O₇ requires 668), 236 (100, C₁₃H₁₈NO₃, trimethoxylated isoquinoline fragment), and 206 (91, 3).

O,O-Diethylthalarugine (22). A 200-mg sample of 19 in 5 mL of MeOH was treated with excess diazoethane generated from 2 g of *N*-ethyl-*N'*-nitro-*N*-nitrosoguanidine. After 5 days the reaction residue after solvent evaporation was chromatographed on 10 g of neutral alumina with 100 mL each of PhH, PhH-CHCl₃ (1:1), and CHCl₃ as eluents. A yield of 120 mg of a pale-yellow amorphous solid of diethyl ether 22 was obtained: R_f 0.85 on TLC with PhH-Me₂CO-NH₄OH (20:20:0.6); NMR (CDCl₃) δ 1.34 (t, $J = 7$, 2 OCH₂CH₃), 3.82 and 4.05 (2 q, $J = 7$, 2 OCH₂CH₃), 2.47 and 2.49 (2 s, 2 NMe), 3.57, 3.81 (2 s, 2 OMe), and 3.78 (s, 2 OMe), 5.91 (s, H-8'), 6.16 (s, H-8), 6.51 (s, H-5), and 6.6-7.1 (m, 7 ArH); MS EI m/e 696 (0.3%, M⁺, C₄₂H₅₂N₂O₇ requires 696), 476 (0.4, M - 8), 446 (1, M - 23), 250 (45, 23), 220 (63, 8), and 192 (100, 8 - CH₂CH₂), with CI

(isobutane) m/e 697 (14, MH⁺), 250 (85, 23) and 220 (100, 8).

Anal. Calcd for C₄₂H₅₂N₂O₇·2H₂O: C, 68.83; H, 7.70; N, 3.82. Found: C, 68.98; H, 7.15; N, 3.95.

Na/NH₃ Cleavage of O,O-Diethylthalarugine (22). A 115-mg sample of 22 in 10 mL of tetrahydrofuran was added dropwise over 1 h to 25 mL of NH₃ containing Na. After an additional 2 h of reaction the products were separated into the phenolic and nonphenolic components by partitioning between Et₂O and 1 N NaOH.

From the ether fraction, 75 mg of a pale-yellow oil, R_f 0.85 on TLC with PhH-Me₂CO-NH₄OH (20:20:0.6), was obtained. Chromatography on 4 g of silica gel with CHCl₃ and 1% MeOH in CHCl₃ gave the nonphenolic base 24 (47 mg), identical (UV, IR, NMR, MS, and CD) with a cleavage product from *O*-ethylthalarugosidine.⁵

The NaOH extract after treatment with solid NH₄Cl and Et₂O extraction gave the phenolic base fraction (35 mg). Chromatography on silica gel with 2 and 3% MeOH in CHCl₃ gave from the latter eluates 23 mg of product 18 that crystallized from MeOH, mp 114 °C, with R_f 0.6 on TLC using the same solvent as for 24. This substance was identical (mmp, UV, IR, NMR, MS, and CD) with a cleavage product from *O*-ethylthalogosine (16).

KMnO₄ Oxidation of O,O-Diethylthalarugine (22). A 315-mg sample of 22 dissolved in 50 mL of Me₂CO was treated with 600 mg of KMnO₄ added portionwise in 1 h while stirring. After an additional 3 h of stirring the excess reagent was decomposed with MeOH, and the MnO₂ was removed by filtration. The filtrate was concentrated, mixed with 25 mL of H₂O, acidified with 5% HCl, and exhaustively extracted with CHCl₃. The residue (210 mg) from the washed (H₂O) and dried (Na₂SO₄) CHCl₃ extract was chromatographed on 10 g of silica gel with 50 mL of PhH and 100 mL each of PhH-CHCl₃ (1:1), CHCl₃, and 2.5, 5, and 10% MeOH in CHCl₃.

The PhH-CHCl₃ (1:1) eluates gave first, after crystallization from MeOH, 36 mg of yellow needles of isoquinolone 25: mp 120-121 °C; R_f 0.72 on TLC with PhH-Me₂CO (1:1); UV, IR, NMR, and MS identical with those of a synthetic sample⁹ of 6-methoxy-7-ethoxy-*N*-methyl-1-oxo-1,2,3,4-tetrahydroisoquinoline.¹⁰

The later PhH-CHCl₃ (1:1) eluates gave 30 mg of isoquinolone 26 as a pale-yellow oil: R_f 0.66 on TLC with the same solvent as used for 25; UV λ_{max} 296 (log ϵ 3.48), 270 (shld) (3.88), 260 (3.96), 253 (shld) (3.92), and 216 nm (4.52); IR (CHCl₃) 1645 cm⁻¹ (C=O); NMR (CDCl₃) δ 1.35 (t, $J = 7$, OCH₂CH₃), 4.06 (q, $J = 7$, OCH₂CH₃), 2.92 and 3.51 (2 A₂B₂ t, $J = 6$, CH₂CH₂), 3.12 (s, NMe), 3.90 (s, 2 OMe), and 7.44 (s, H-8); MS m/e 265.1320 (100%, M⁺, C₁₄H₁₉NO₄ requires 265.1314), 250 (3, M - Me), 236 (9, M - Et), 222 (33, M - CH₂NMe), and 194 (35, 222 - CO). This compound was identical (TLC, UV, IR, NMR, and MS) with 5-ethoxy-6,7-dimethoxy-*N*-methyl-1-oxo-1,2,3,4-tetrahydroisoquinoline produced by KMnO₄ oxidation of the *O*-ethylated cleavage product 10 from authentic thalidasine (5).¹¹

From the 5 and 10% MeOH in CHCl₃ fractions, 100 mg of a white solid was obtained that exhibited the same TLC characteristics as 2-methoxy-4',5'-dicarboxy diphenyl ether (27) prepared from the corresponding dialdehyde¹² by Ag₂O oxidation. Treatment of both samples with diazomethane afforded diester 28, mp 79-80 °C, with identical TLC mobility, IR and NMR spectra, and an undepressed mmp.

Preparation of O,O-Diethyl 10 and KMnO₄ Oxidation. Diphenol 10 (120 mg) obtained by Na/NH₃ cleavage¹¹ of thalidasine (5, 300 mg) in 5 mL of MeOH was treated with excess ethereal diazoethane. After 12 days the solvent was evaporated and the residue chromatographed on 5 g of neutral alumina with PhH (200 mL) and PhH-CHCl₃ (4:1, 100 mL). From the latter eluate, 80 mg of the diethyl ether of 10 as a yellow heavy oil was obtained: R_f 0.86 on TLC with PhH-Me₂CO-NH₄OH (10:10:0.1); UV λ_{max} 278 nm (log ϵ 3.77) and unchanged by acid or base; NMR (CDCl₃) δ 1.34 and 1.38 (2 t, $J = 7$, 2 OCH₂CH₃), 3.98 and 4.05 (2 q, $J = 7$, 2 OCH₂CH₃), 2.50 (s, NMe), 3.57 and 3.82 (2 s, 2 OMe), 5.91 (s, H-8), and 6.77 and 6.97 (AA'BB' q, $J_{AB} = 9$, 4 ArH); MS m/e 385 (0.2%, M⁺), 250 (100, 23), 221 (1, 23 - Et), 220 (8, 23 - Et-H) and 135 (2, M - 23).

A 70-mg sample of *O,O*-diethyl-10 was dissolved in 25 mL of Me₂CO, and 150 mg of KMnO₄ was added during 1 h while stirring. After reacting an additional 3 h the excess reagent was destroyed by 10 mL of MeOH. The mixture was filtered and the filtrate concentrated to 3 mL. Addition of 10 mL of H₂O and acidification with 0.1 N HCl produced a white precipitate that was removed by filtration. The filtrate was extracted with 150 mL of CHCl₃, and the extract after washing (H₂O) and drying (Na₂SO₄) left, on evaporation, 40 mg of a pale-yellow oil. It was chromatographed on 2 g of neutral alumina with PhH (50 mL). The 30 mg of heavy oil was identified by spectral evidence to be isoquinolone 26, identical (TLC, UV, IR, NMR, and MS) with the corresponding oxidation products from *O,O*-diethylthalarugine (22) and *O,O*-diethylthalarugidine (34).

Thaliruginine (29). From the same column separation that gave thaligosine (14), the 3 and 4% MeOH in CHCl_3 eluates gave 260 mg of thaliruginine (29) as a white amorphous solid: R_f 0.65 on TLC with $\text{PhH-Me}_2\text{CO-NH}_4\text{OH}$ (20:20:0.8); $[\alpha]_D^{20} +104^\circ$ (c 0.16, MeOH); CD $[\theta]_{287} +14\ 500$, $[\theta]_{252} -2100$, $[\theta]_{230} +93\ 000$; UV λ_{max} 281 nm ($\log \epsilon$ 3.90) and in 0.01 N NaOH 281 (3.97) and 309 nm (shld) (3.09); IR (CHCl_3) $3520\ \text{cm}^{-1}$ (OH); NMR (CDCl_3) δ 2.48 and 2.50 (2 s, 2 NMe), 3.57, 3.61, 3.78, 3.80 and 3.83 (5 s, 5 OMe), 5.71 (s, H-8), 6.11 (s, H-8'), 6.53 (s, H-5'), 6.6-7.2 (m, 7 ArH), and 5.4 (br, OH, D_2O exchangeable); MS m/e 654 (0.8%, M^+), 222 (68, 20), 206 (100, 3), and 192 (26, 3 - Me). Thaliruginine gave positive tests with phosphomolybdic acid⁷ and Gibbs' reagent.⁸

Anal. Calcd for $\text{C}_{39}\text{H}_{46}\text{N}_2\text{O}_7\cdot\text{H}_2\text{O}$: C, 69.62; H, 7.19. Found: C, 69.32; H, 6.86.

O-Methylation of Thaliruginine (29) to 21. A 40-mg sample of 29 in 5 mL of MeOH was treated with excess ethereal diazomethane for 3 days to give 35 mg of derivative 21, identical (TLC, UV, IR, NMR, and CD) with the methylated product of thalirugine (19).

O-Ethylthaliruginine (30). A 100-mg sample of 29 in 5 mL of MeOH was treated with excess ethereal diazoethane. The product was chromatographed on 5 g of silica gel PF 254 using 100 mL each of CHCl_3 and 1% MeOH in CHCl_3 as eluents to yield 80 mg of 30 as an amorphous pale-yellow solid: R_f 0.8 on TLC with $\text{PhH-Me}_2\text{CO-NH}_4\text{OH}$ (20:20:0.8); NMR (CDCl_3) δ 1.34 (t, $J = 7$, OCH_2CH_3), 4.05 (q, $J = 7$, OCH_2CH_3), 2.48 and 2.50 (2 s, 2 NMe), 3.58, 3.62, 3.78, 3.80, and 3.82 (5 s, 5 OMe), 5.92 (s, H-8'), 6.13 (s, H-8), 6.58 (s, H-5), and 6.6-7.2 (m, 7 ArH); MS m/e 682 (0.2%, M^+ , $\text{C}_{41}\text{H}_{50}\text{N}_2\text{O}_7$ requires 682), 250 (95, 23), and 206 (100, 3).

Na/NH₃ Cleavage of O-Ethylthaliruginine (30). A 60-mg sample in 5 mL of tetrahydrofuran was reacted with Na/NH₃, and the products were worked up as described for compound 22. From the nonphenolic fraction, 11 mg of (*S*)-*O*-methylarmepavine (31) was obtained and identified by direct comparison (TLC, UV, IR, NMR, and CD) with an authentic sample. The phenolic fraction gave 10 mg of crystalline 18 which was identified by comparison (mmp, TLC, UV, IR, NMR, and CD) with an authentic sample.

Thalirugidine (32). The residue (360 mg) from column fraction 37 was rechromatographed on 16 g of silica gel with CHCl_3 and 2, 4, 8, and 10% MeOH in CHCl_3 as eluents. From the 2% MeOH in CHCl_3 eluate, 55 mg of thalirugidine (32) was obtained as a white amorphous solid: R_f 0.60 on TLC with $\text{PhH-Me}_2\text{CO-NH}_4\text{OH}$ (20:20:0.8); $[\alpha]_D^{20} +112^\circ$ (c 0.19, MeOH); CD $[\theta]_{280} +7200$, $[\theta]_{230} +98\ 500$; UV λ_{max} 278 nm ($\log \epsilon$ 3.82) and in 0.01 N NaOH 281 nm (3.94); IR (CHCl_3) $3530\ \text{cm}^{-1}$; NMR (CDCl_3) δ 2.48 and 2.51 (2 s, 2 NMe), 3.61, 3.63, 3.85 (3 s, 3 OMe) and 3.81 (s, 2 OMe), 5.76 and 5.79 (2 s, H-8 and H-8'), 6.6-7.2 (m, 7 ArH), and 5.1 (br, 2 OH, D_2O exchangeable); MS m/e 670 (1.4%, M^+), 222 (100, 20), 221 (3, 20 - H), 220 (3, 20 - 2H), 207 (3, 20 - Me), 206 (8, 20 - Me - H), 192 (8, 220 - CO), and 178 (2, 206 - CO).

Anal. Calcd for $\text{C}_{36}\text{H}_{46}\text{N}_2\text{O}_8$: C, 69.83; H, 6.91; N, 4.18. Found: C, 69.78; H, 7.18; N, 4.15.

O, O-Dimethylthalirugidine (33). Thalirugidine (32, 20 mg) in 2 mL of MeOH was treated with excess diazomethane for 3 days to give 15 mg of a pale-yellow amorphous solid; R_f 0.83 on TLC with $\text{PhH-Me}_2\text{CO-NH}_4\text{OH}$ (20:20:0.8); $[\alpha]_D^{20} +53^\circ$ (c 0.21, MeOH); CD $[\theta]_{285} +8700$, $[\theta]_{230} +42\ 000$; UV λ_{max} 278 nm ($\log \epsilon$ 3.96); NMR (CDCl_3) δ 2.47 and 2.50 (2 s, 2 NMe), 3.59, 3.61, 3.79 (3 s, 3 OMe) and 3.83 (s, 4 OMe), 5.93 and 5.98 (2 s, H-8 and H-8'), and 6.6-7.2 (m, 7 ArH); MS m/e (M^+ not observed), 236 (100, isoquinoline fragments).

O, O-Diethylthalirugidine (34). A 130-mg sample of 32 in 5 mL of MeOH was treated with diazoethane for 5 days. The reaction residue was chromatographed on 6 g of silica gel using 1% MeOH in CHCl_3 as eluent, to give 120 mg of the diethyl ether 34 as an amorphous solid: $[\alpha]_D^{20} +44^\circ$ (c 0.17, MeOH); CD $[\theta]_{282} +5700$, $[\theta]_{230} +30\ 500$; NMR (CDCl_3) δ 1.33 (t, $J = 7$, $2\ \text{CH}_2\text{CH}_3$), 4.03 (q, $J = 7$, $2\ \text{CH}_2\text{CH}_3$), 2.45 and 2.47 (2 s, 2 NMe), 3.58, 3.60, 3.77 (3 s, 3 OMe), 3.81 (s, 2 OMe), 5.92 and 5.96 (2 s, H-8 and H-8'), and 6.6-7.2 (m, 7 ArH); MS m/e 726 (0.5%, M^+ , $\text{C}_{43}\text{H}_{54}\text{N}_2\text{O}_8$ requires 726), 250 (100, 23), 235 (8, 23 - Me), 234 (7, 23 - Me - H), 221 (4, 23 - Et), 220 (17, 23 - Et

- H), 206 (11, 234 - CO), and 192 (7, 220 - CO).

Na/NH₃ Cleavage of O, O-Diethylthalirugidine (34). The diethyl ether 34 (105 mg) in 10 mL of tetrahydrofuran was reductively cleaved with 150 mg of Na in ammonia, and the products were divided into phenolic and nonphenolic bases. The nonphenolic fraction (50 mg) as a yellow oil showed two spots, R_f 0.86 (major) and 0.91, on TLC with $\text{PhH-Me}_2\text{CO-NH}_4\text{OH}$ (20:20:0.8). Chromatography on 2.5 g of silica gel PF 254 with 50 mL of PhH-CHCl_3 (1:1) and 100 mL of CHCl_3 gave 18 mg of 35, the major base, as a colorless oil. It showed TLC mobility and UV, IR, NMR, and CD spectra identical with the corresponding cleavage product from thalidezine¹³ or isothalidezine.¹⁴ The minor product 36, obtained as a yellow oil (8 mg), showed identical TLC characteristics and UV, IR, and NMR spectra as those for a cleavage by-product from thalidezine.¹³

The phenolic fraction (38 mg) contained one component, R_f 0.36 on TLC with system used for the nonphenolics. Chromatography on 2 g of silica gel PF 254 with 100 mL each of 1 and 2% MeOH in CHCl_3 afforded a crystalline residue (26 mg) that was recrystallized with MeOH to give colorless rosettes of 18, mp 113-114 °C, identical (TLC, UV, IR, NMR, and CD) with a sample from *O*-ethylthaligosine (16).

Methylation of 18 with diazomethane gave 35.

KMnO₄ Oxidation of O, O-Diethylthalirugidine (34). Compound 34 (50 mg) in 10 mL of Me_2CO was treated with 150 mg of KMnO_4 . After stirring for 3 h, the workup as reported for the oxidation of 22 yielded 8 mg of isoquinolone 26 and the diphenyl ether diacid 27. Both were compared directly (the latter as the dimethyl ester 28) with known samples by TLC and UV, IR, and NMR spectra. Compound 28 also showed no depression of the mmp.

Acknowledgment. We thank the National Institutes of Health, United States Public Health Service, for the grant (HL-07502) supporting this work and Mr. R. Weisenberger of the Chemistry Department for some of the mass spectra.

Registry No.—1, 64252-82-0; 6, 64252-83-1; 9, 3423-12-9; 10, 16623-60-2; 10 diethyl ether, 64235-37-6; 11, 64235-38-7; 14, 22226-72-8; 16, 64235-39-8; 18, 64235-40-1; 19, 64235-41-2; 21, 64215-90-3; 22, 64215-91-4; 25, 2651-56-1; 26, 64215-92-5; 28, 5566-15-4; 29, 64215-93-6; 30, 64215-94-7; 32, 64215-95-8; 33, 64215-96-9; 34, 64215-97-0; 35, 64281-58-9; 36, 64281-59-0.

References and Notes

- Alkaloids of *Thalictrum* 25. For paper 24 see: J. Wu, J. L. Beal, W.-N. Wu, and R. W. Doskotch, *Lloydia*, in press.
- For reviews up to 1970, see: (a) P. L. Schiff, Jr., and R. W. Doskotch, *Lloydia*, **33**, 403 (1970); (b) N. M. Mollov and V. St. Georgiev, *Recent Dev. Chem. Nat. Carbon Comp.*, 195, 257 and 301 (1971); N. M. Mollov and H. B. Dutschewska, *ibid.*, 202 (1971).
- See the following and references therein: (a) W.-N. Wu, J. L. Beal, G. W. Clark, and L. A. Mitscher, *Lloydia*, **39**, 65 (1976); (b) N. M. Mollov, Le Nhat Thuan, and P. P. Panov, *Dokl. Bolg. Akad. Nauk*, **24**, 1047 (1971); (c) N. M. Mollov, I. C. Ivanov, V. St. Georgiev, P. P. Panov, and N. Kotsev, *Planta Med.*, **19**, 10 (1971); (d) T. Cieszynski and B. Borowski, *Acta Pol. Pharm.*, **22**, 440 (1965).
- J. Baldas, I. R. C. Bick, T. Ibuka, R. S. Kapil, and Q. N. Porter, *J. Chem. Soc., Perkin Trans. 1*, 597 (1972), and references therein.
- L. A. Mitscher, W.-N. Wu, R. W. Doskotch, and J. L. Beal, *Lloydia*, **35**, 167 (1972).
- T. Kugo, *Yakugaku Zasshi*, **79**, 322 (1959).
- V. M. Platkovska and S. G. Vatkina, *J. Appl. Chem. (USSR)*, **10**, 202 (1937).
- F. E. King, T. J. King, and L. C. Manning, *J. Chem. Soc.*, 563 (1957).
- Prepared by Dr. W.-T. Liao from synthetic thalifoline by treatment with diazoethane. Thalifoline was made according to R. W. Doskotch, P. L. Schiff, Jr., and J. L. Beal, *Tetrahedron*, **25**, 469 (1969).
- E. Späth and H. Epstein, *Chem. Ber.*, **59**, 2791 (1926).
- S. M. Kupchan, T.-H. Yang, G. S. Vasilikiotis, M. H. Barnes, and M. L. King, *J. Org. Chem.*, **34**, 3884 (1969).
- T. Kametani and K. Fukumoto, *J. Chem. Soc.*, 6141 (1964).
- M. Shamma, R. J. Shine, and B. S. Dudock, *Tetrahedron*, **23**, 2887 (1967).
- W.-N. Wu, J. L. Beal, R.-p. Leu, and R. W. Doskotch, *Lloydia*, **40**, 384 (1977).